

UNCLASSIFIED

| |
|--|
| |
| |
| |
| AD NUMBER |
| AD485850 |
| NEW LIMITATION CHANGE |
| TO Approved for public release, distribution unlimited |
| FROM Distribution authorized to U.S. Gov't. agencies and their contractors; Administrative/Operational Use; Jun 1966. Other requests shall be referred to US Army Biological Center, Fort Detrick, Frederick, MD 21701. |
| AUTHORITY |
| BDRL, D/A ltr dtd 28 Sep 1971 |

THIS PAGE IS UNCLASSIFIED

48850

AD

TECHNICAL MANUSCRIPT 294

PROPERTIES OF VENEZUELAN
EQUINE ENCEPHALOMYELITIS VIRUS
ACCOMPANYING ATTENUATION IN VITRO

Henry J. Hearn, Jr.
William T. Soper

JUNE 1966

Best Available Copy

UNITED STATES ARMY
BIOLOGICAL CENTER
FORT DETRICK

Reproduction of this publication in whole or part is prohibited except with permission of Commanding Officer, U. S. Army Biological Center, ATTN: Technical Releases Group, Technical Information Department, Fort Detrick, Frederick, Maryland, 21701. However, DDC is authorized to reproduce the publication for United States Government purposes.

DDC AVAILABILITY NOTICES

Qualified requesters may obtain copies of this publication from DDC.

Foreign announcement and dissemination of this publication by DDC is not authorized.

Release or announcement to the public is not authorized.

DISPOSITION INSTRUCTIONS

Destroy this publication when it is no longer needed. Do not return it to the originator.

The findings in this publication are not to be construed as an official Department of the Army position, unless so designated by other authorized documents.

Best Available Copy

U.S. ARMY BIOLOGICAL CENTER
Fort Detrick, Frederick, Maryland

TECHNICAL MANUSCRIPT 294

PROPERTIES OF VENEZUELAN EQUINE ENCEPHALOMYELITIS VIRUS
ACCOMPANYING ATTENUATION IN VITRO

Henry J. Hearn, Jr.

William T. Soper

Virus and Rickettsia Department
BIOLOGICAL SCIENCES LABORATORY

Project 1C014501B71A

June 1966

In conducting the research reported here, the investigators adhered to "Principles of Laboratory Animal Care" as established by the National Society for Medical Research.

ABSTRACT

Virus obtained during serial plaque passage of the virulent parent egg seed (PES) strain of Venezuelan equine encephalomyelitis (VEE) virus produced only large plaques during either three serial plaque passages in chick fibroblasts or ten plaque passages in L cells and was lethal for mice by the intraperitoneal (IP) route. Virus showing these characteristics was designated the stable large-plaque (Ls) type. In contrast, virus obtained during serial plaque passage of the attenuated 9t strain in chick fibroblasts formed only very small plaques and was not lethal for mice by the IP route. Virus showing these properties was designated the stable small-plaque (Ss) type. Under other passage conditions, however, large-plaque virus that yielded about 90% large and 10% small plaques was obtained; this virus was designated the unstable large or Lu type because it differed from the Ls type, which yielded only large plaques. The Lu type continued to yield the same ratio of large to small plaques for several plaque-to-plaque passages. In addition, small-plaque virus that yielded both large and small plaques and that showed a reduced capability to infect mice was also recovered. This virus was designated the unstable small or Su type because it differed from the Ss type in its higher level of virulence and in its plaque-forming properties. Thus, based upon the properties of virulence for mice and plaque size, four viral types could be discerned. The evidence suggests that serial passage in cell culture imposed environmental pressures that sequentially selected the following viral types: Ls, Lu, Su, and Ss.

I. INTRODUCTION

Several Venezuelan equine encephalomyelitis (VEE) virus variants have been described that showed a marked loss in virulence for common laboratory animals after passage in cell cultures.¹⁻⁵ Perhaps the most significant of these is the strain of Berge et al.,⁴ whose variant appears to be suitable as a live vaccine for humans.⁶ Studies with a VEE virus variant isolated in this laboratory³ and with the viral strain described by Musgay and Suarez⁵ showed that the loss in virulence of the virus was attended by a loss in ability to form large plaques.^{5,7} Brown⁸ reported that a high-passage preparation of the avirulent strain isolated by Berge and his colleagues apparently produced only small plaques. Heydrick et al.⁹ showed that during ten serial passages of VEE virus in L cells or chick fibroblasts, plaque size began to decrease as attenuation became demonstrable. Detailed information, however, concerning mutational events that may occur during the conversion to small-plaque attenuated VEE virus strains is lacking. In this report, an attempt was made to recognize and to define some of the mutational stages that occurred during the attenuation of our strains of VEE virus by passage in vitro. Viral populations that arose under various conditions of selection and passage were studied and characterized.

II. MATERIALS AND METHODS

A. VIRUS STRAINS

The parent egg seed strain, described elsewhere,³ was obtained originally from the brain of a donkey that had succumbed to VEE virus disease. The virus subsequently received 13 passages in embryonated eggs prior to its use as the stock parent egg seed (PES).

The attenuated virus (9t) strain was obtained from an L cell monolayer culture that had become chronically infected after inoculation of the PES strain. The virus was harvested 8 months after initial infection and underwent a number of serial passages in fresh L cells. The virus from the ninth such serial passage was designated the 9t strain. Properties of this strain have been described elsewhere.³

B. CELL CULTURES FOR GROWTH AND PLAQUE ASSAY

Propagation and plaque assays of virus were performed in chick fibroblast and, when indicated, in L cell monolayers. Chick fibroblasts were prepared by a modification of the procedure described by Dulbecco.¹⁰ They were grown in Medium 199 supplemented with 20% horse serum plus 0.1 mg streptomycin and 100 units penicillin per milliliter. Confluent cell sheets were routinely obtained in 30-mm plastic plates after 48 hours of incubation at 37 C in an atmosphere of 5% carbon dioxide. L cell cultures were obtained from stock cultures at these laboratories and grown in a similar manner.

Virus suspensions to be assayed were routinely diluted in 0.1% yeast extract - 0.1% proteose peptone #3 in Hanks BSS (pH 7.4) and appropriate concentrations were inoculated directly onto the cell sheets to provide counts of five to fifty plaques ranging over two tenfold dilutions. After a 30-minute adsorption period the excess inoculum was removed and 5 ml of agar overlay was added. The overlay consisted of lactalbumin hydrolyzate (0.45%), yeast extract (0.09%), Bacto-agar (Difco) (1.1%), and 0.5% gelatin. Infected monolayers were incubated at 37 C with 5% CO₂ for 72 hours and stained by adding a second agar overlay containing 1:10,000 neutral red. Plaques were visible within 3 hours after staining.

Virus from individual plaques was isolated with a capillary pipette that removed agar, virus, and a portion of the cell sheet within the plaque. This was dispersed in 2.5 ml of yeast extract - peptone diluent, containing 50% horse serum, and was considered as undiluted plaque-virus. Generally, such isolates were stored at -55 C before use. Virus was then inoculated directly onto fresh chick fibroblasts or L cells for plaque assay and for tests for virulence in mice. In some instances, because of the small quantity of virus present in some plaques, isolates were grown in fluid cultures of chick fibroblasts for 48 to 72 hours to increase the virus concentration prior to the plaque assays and inoculations in mice. Titers of such viral suspensions were 10⁶ to 10⁷ MICLD₅₀ per ml.

C. VIRUS TITRATIONS

Virus samples were titrated intracerebrally (IC) and intraperitoneally (IP) in 12- to 14-g Swiss white mice and the titers expressed as MICLD₅₀/ml and MIPLD₅₀/ml, respectively. In most instances, particularly with virus from single plaques, titration was not necessary and an appraisal of virulence was obtained by injecting mice with undiluted and/or 1:10 dilutions of virus. Mice that survived injections with the plaque material were challenged by the IP route after 21 days with a multiple lethal dose (10^{3.5} to 10^{4.5} MICLD₅₀) of the PES strain. Mice that were immune to challenge were considered as having been subclinically infected as a result of the initial injection.

III. RESULTS

A. CHARACTERIZATION OF THE VIRULENT PARENT (PES) AND ATTENUATED (9t) VEE VIRUS STRAINS

Plaques ordinarily formed by the PES strain during first passage in chick fibroblasts ranged from 3 to 6 mm in diameter; the majority were 4 mm. In all of our experiments, examining many hundreds of plaques, diameters smaller than 3 mm were rarely observed. Although the PES strain possesses limited virulence in monkeys, eliciting chiefly a febrile, non-lethal response, this strain is uniformly lethal in mice and other laboratory animals when injected IP. Results of tests for virulence and plaque size, performed with virus obtained during the second and third plaque passages, were the same as those during the first passage. During the fourth, sixth, and ninth serial passage of large-plaque virus in chick fibroblasts, however, both large and small (2 mm) plaques were found; when present, the latter amounted to 8 to 10% of the total plaque number. Results of tests during ten serial passages of large-plaque virus in L cells were similar to those found during three serial passages in chick fibroblasts; small-plaque virus was rarely observed.

Virus from the PES strain that formed only large plaques during either three passages in chick fibroblasts or ten passages in L cells appeared homogeneous and genetically stable. Such virus was designated as a stable, large-plaque (Ls) type. Large-plaque virus that yielded mixtures of both large and small plaques beyond the third serial plaque passage in chick fibroblasts was designated as an unstable, large-plaque (Lu) type. Virus isolated from the large plaques in either cell type was lethal for mice by the IP route. A more detailed characterization of Lu plaque types isolated from the PES strain is presented in a later section of this report.

In contrast to results with the PES strain, the 9t strain formed small plaques (0.5 to 1.5 mm) in chick fibroblasts. Except for a few deaths found occasionally with undiluted virus, the strain was nonlethal for mice by the IP route.

The virus isolated from single, small plaques formed only small plaques containing attenuated virus during either four plaque passages in chick fibroblasts or ten passages in L cells. For this reason, virus of this type was considered, by definition, to be relatively homogeneous and genetically stable. It was designated as a stable small-plaque (Ss) type.

B. DERIVATION OF UNSTABLE LARGE-PLAQUE (Lu) TYPES FROM THE 9t STRAIN

In one experiment the virus from six small plaques was pooled after the fourth serial passage of small-plaque virus from the 9t strain. This preparation yielded plaques of which 2 (4%) were 0.5 to 1.5 mm and 44 (96%) were 2 to 3 mm in diameter. On the same plate one 7-mm plaque was found. The emergence of plaques of the 2- to 3-mm range from small-plaque virus of the attenuated strain possibly represented a rare instance in which plaques were 1- to 2-mm larger than those usually observed. This observation might have resulted from undetermined environmental changes that temporarily influenced plaque size only. Another explanation for the appearance of the 2- to 3-mm plaques is that they might have been intermediate between the large type usually produced by the parent (PES) strain and the small attenuated (9t) type with respect to size and perhaps to virulence. In the absence of immediately suitable evidence to the contrary and for purposes of classification, these plaques were tentatively considered small plaques. The 7-mm plaque, however, contained particles that were lethal for mice when injected IP. Virus isolated from this plaque was selected for further study.

The data in Figure 1 show that the virus isolated from the large, 7-mm plaque formed 87% large and 13% small plaques. This virus resembled the stable, large-plaque (Ls) virus found in the PES strain because it formed large plaques and was lethal for mice. It differed from the Ls type, however, because it was unstable and produced a high percentage of small plaques in addition to the large plaques upon passage. For this reason, this virus was considered as another example of the unstable large-plaque (Lu) type. The results of the passage of virus from two 5-mm plaques that came from the 7-mm plaque also are represented in Figure 1. The virus from one of these, designated plaque 5a, produced 94% large and 6% small plaques (shown on Line A). Deliberate selection and passage of the virus from large plaques continued to result in the formation of approximately 90% large and 10% small plaques. Virus recovered from the large plaques was always virulent for mice.

The results suggest that the virus from plaque 5a and its progeny was genetically unstable and indistinguishable from that of the parent 7-mm plaque. Large-plaque isolates from this line, therefore, continued to warrant the Lu designation.

In contrast, the virus isolated from plaque 5b immediately produced 100% small plaques during the same experiment in which the results from the passage of virus in plaque 5a were obtained. Figure 1 (Line B) shows that two passages of this small-plaque virus resulted in the continued formation of 100% small plaques. A further passage of virus from a pool of four plaques again resulted in 100% small plaques. Comparable results were obtained for eight additional passages, and mice that were injected with the avirulent small-plaque virus were resistant to challenge with the lethal PES strain.

Figure 1 (top of Line B) also shows that the virus obtained from a 2-mm plaque, and passed at the same time as the virus described above, contained only virus that yielded small (1-mm) plaques. Virus from a pool of five small plaques was serially passed twice in chick fibroblast fluid cultures to provide an opportunity for large-plaque, virulent virus to emerge. Once again, virus in the supernatant culture fluid produced only small plaques and was not lethal for mice.

The results obtained with the virus from plaque 5b suggest that it represented an Lu type that may have contained a very small percentage of large-plaque virus. Passage of the 5b plaque isolate gave rise to a stable, small-plaque (Ss) type, the details of which are discussed further in the following section.

C. DERIVATION OF UNSTABLE SMALL-PLAQUE (Su) AND STABLE SMALL-PLAQUE (Ss) TYPES FROM UNSTABLE LARGE-PLAQUE (Lu) TYPES

Results of attempts to examine more critically the progeny obtained after passage of virus from Lu plaque types are presented below and in Figure 2. These data indicate that virus from Lu plaques gave rise to an unstable small (Su) variety that, in turn, formed stable small-plaque (Ss) types.

As one example of this, virus from plaque 5a was serially passed three times in chick fibroblast fluid cultures in order to provide suitable quantities of working material. The results are shown in Figure 2, Line A. Viral material from the third serial passage formed 63% large and 37% small plaques; it was lethal for two of six mice and induced paralytic illness in two others that eventually recovered. Continuing along Line A, virus isolated from a small plaque (1.5 mm) formed 33% large and 67% small plaques. Because the virus from this plaque contained significant quantities of both large- and small-plaque-forming particles, it was considered to be an unstable, small-plaque (Su) type. After the virus was passed in chick fibroblast fluid cultures, however, it produced 100% plaques of the Ss type and was nonlethal for mice. Nonlethal infectivity of the Ss virus in mice was demonstrated by the resistance of the mice to a challenge with a multiple lethal dose of the PES strain.

Under another set of circumstances (Figure 2, Line B), the virus from plaque 5a was serially passed only twice in chick fibroblast fluid cultures. Virus obtained 48 hours postinoculation during the second passage produced only large plaques. Virus obtained at 72 hours from the same cultures (Line C), however, produced 82% large and 18% small plaques; this virus was lethal for mice. The opportunity for additional cycles of viral growth from 48 to 72 hours in the chick fibroblasts apparently led to the appearance of small quantities of small-plaque virus. Continuing along Line C, virus from a 1.5-mm plaque, representative of the small plaques on the plate, was passed in chick fibroblast fluid cultures to increase the titer and then subjected to a plaque test.

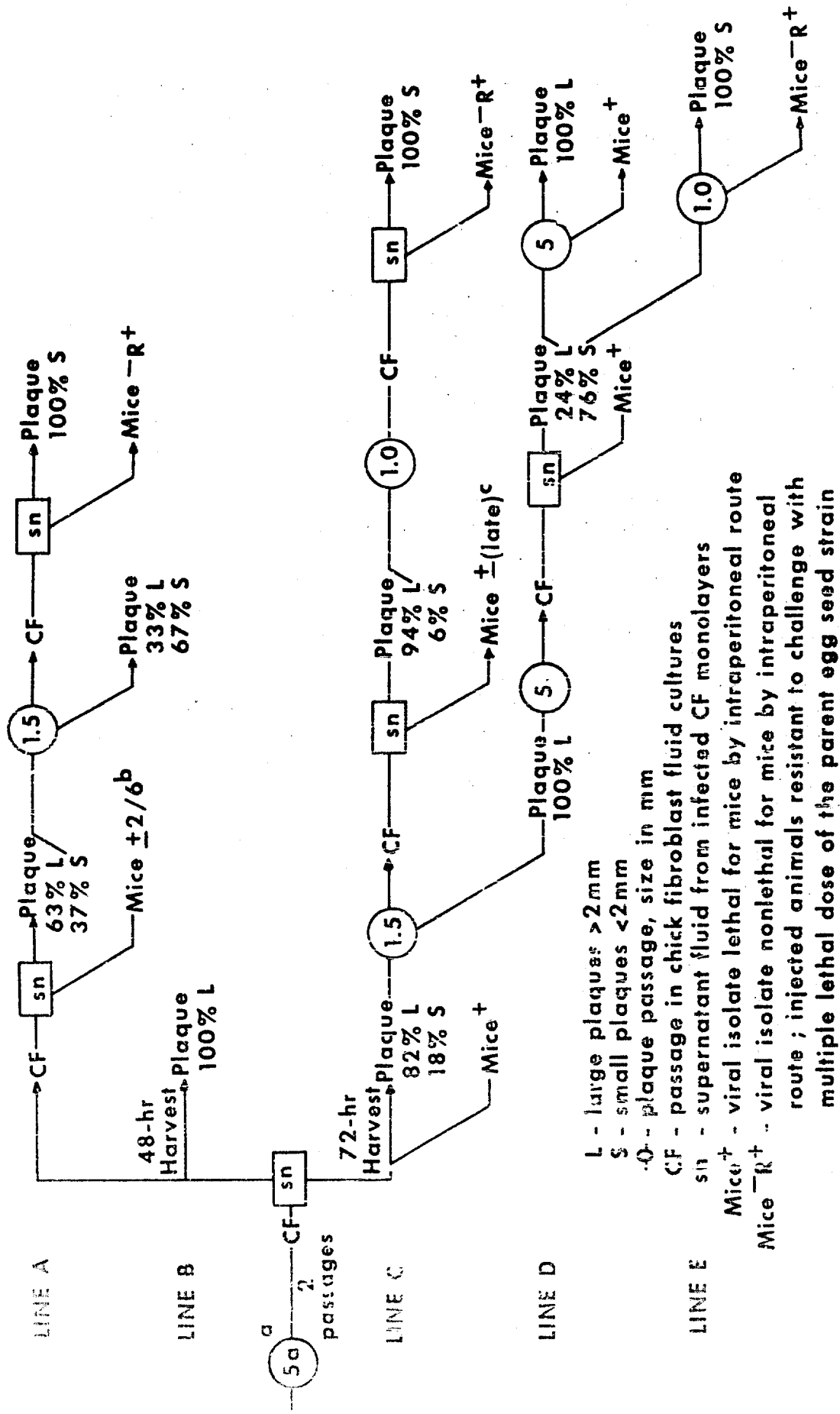


Figure 2. Derivation of Large Unstable, Small Unstable, and Small Stable Viral Plaque Types from a Large Unstable Viral Plaque Type.

a. Same as plaque 5a in Figure 1.

b. Virus lethal for two of six mice, two other paralyzed mice recovered.

c. Mean day of death delayed 48 hours.

After passage of the 1.5-mm plaque virus in fluid culture (Line C), the harvested material produced 94% large plaques, 6% small plaques, and was lethal for mice. Thus, the virus from this (1.5-mm) plaque was not typical of stable, small-plaque virus of the Ss type because it contained virus capable of forming both large and small plaques and producing lethal infections in mice by the IP route. It is of interest that mice that succumbed to this virus did so 2 to 4 days beyond the usual course of the disease. For these reasons, this atypical, small-plaque virus from the 1.5-mm plaque was considered to be another example of the unstable small (Su) type. From the resultant 6% small plaques, 1-mm plaque virus was isolated and grown in chick fibroblast fluid cultures. Supernatant fluids from these cultures contained 100% small-plaque virus. This virus appeared to be similar, if not identical, to virus of the Ss-type. That the mice were sublethally infected by Ss type virus was shown by the fact that they resisted a challenge with a multiple lethal dose of the PES strain.

Returning to the information in Figure 2, Line D, it can be seen that virus obtained directly from the 1.5-mm Su plaque (shown on Line C), and passed at the same time as the small plaques discussed above, produced only large plaques. Virus from the large plaques appeared to be of the Lu type. As shown on Line D, large plaques were encountered in the minority (e.g., 24% large vs. 76% small) if the large-plaque virus had an intervening passage in chick fibroblast fluid cultures prior to being plaque-tested. These data support the previous observations that the passage of virus of the Lu type in chick fibroblast fluid cultures, allowing multiple cycles of replication, appeared to favor the selection of small-plaque virus. The final results, on Line E (far right side of Figure 2), indicate that small-plaque virus of the Ss type also could be recovered from supernatant fluids of chick fibroblast cultures infected with a 5-mm, Lu plaque type. Typically, this Ss virus formed only small plaques and was nonlethal for mice; animals injected with this virus resisted a later challenge with a multiple dose of the PES strain.

D. DERIVATION OF ATTENUATED, STABLE SMALL-PLAQUE (Ss) TYPES FROM THE VIRULENT, STABLE LARGE-PLAQUE (Ls) TYPE

The foregoing data suggest that passage of virus in chick fibroblasts provided conditions that were selective for stable small-plaque, attenuated virus. Thus far, however, Ss virus was demonstrated as having originated only from large plaques of the Lu type in the 9t strain. This raised the question as to whether the Ss type could be derived from the Ls type, of which the virulent PES strain appears to be almost entirely composed.

It has already been mentioned that three serial passages of PES-strain virus from large plaques resulted in yields of similar plaque types. It was found, however, that by taking advantage of the selective environmental conditions apparently existing during the subsequent serial

plaque passage of virus in chick fibroblasts, certain small-plaque-forming mutants could be recognized and isolated. As an example of this, from among the large plaques formed by the PES virus material, the virus from four 3-mm plaques was pooled. Upon passage of this pool, the majority of plaques was again found to be large (3 to 4 mm) but several were as small as 2 mm. This suggested that the initial step, namely the conversion of virus from the Ls to that of the Lu type, had occurred. Passage of one of the 2-mm plaques resulted in progeny, the majority of which produced plaques approximately 2 mm in size and several as small as 0.5 mm. This suggested the occurrence of the phenomenon similar to that discussed in previous sections, in which the Lu type gave rise to virus of the Su type.

From among the small (<2 mm) plaques, virus from a single 1-mm plaque was passed further, resulting in plaques ranging between 0.5 and 2.0 mm in diameter. This last procedure was repeated to make a total of 10 serial passages from the PES strain starting material. Virus that was harvested from 1-mm plaques had become nonlethal for mice by the IP route at this passage level. Previous to the tenth passage some evidence of virulence for mice by the IP route had persisted. Results during the eleventh and twelfth serial passages demonstrated that stable small-plaque (0.5 to 1 mm), attenuated virus had arisen after ten serial passages of virus from the stable large-plaque, virulent PES strain.

IV. DISCUSSION

The serial passage of various isolates of VEE virus in chick fibroblasts in these studies disclosed the presence of environmental pressures that, in the main, selected for small-plaque, attenuated particles. These viral types could be recognized by their reduced ability to form large plaques under agar and cause lethal illness in mice by the IP route. This provides additional support for related observations on plaque size and virulence that have been made previously with VEE virus^{3,5,7,8} and with poliovirus,¹¹ vesicular exanthema virus of swine,¹² and Coxsackie virus.¹³ It is interesting to note, however, that attenuation has not been found to be associated with small-plaque isolates of Sindbis virus.¹⁴

Less efficient counter-pressures appeared to cause a sporadic recurrence of small numbers of back-mutant large-plaque virulent particle types. This was disclosed initially during our examination of a pool of virus from six small plaques that were obtained from the attenuated 9t strain. Although the large-plaque back-mutants encountered during our experiments superficially resembled the large-plaque virus of the unattenuated PES strain, the virus contained in the large-plaque mutant was, in fact, different. These mutants consistently yielded relatively uniform mixtures

of large and small plaques upon passage under agar whereas large-plaque virus isolated directly from the PES strain yielded large plaques only. Because of this difference, virus from large plaques that yielded both large and small plaques was designated the unstable large or Lu type and the virus from the PES strain was designated the stable large or Ls type. It is of further interest that the small plaques obtained from passage of the large-plaque back-mutant contained small-plaque virus that resembled the attenuated strain in plaque size. This similarity in plaque size also proved to be superficial because the small-plaque virus (Su) isolated from large-plaque back-mutants almost always showed a higher level of virulence than that of the Ss virus from the attenuated strain. In addition, the unstable small (Su) types almost always resulted in mixtures of both large and small plaques upon passage. The latter, Ss type, yielded small plaques only. Continued passage of virus of the Su type resulted in the recovery of Ss type virus typical of that found in the 9t strain.

Thus, based upon the properties of virulence for mice and plaque size, four viral types could be discerned. When the data on the occurrence of these plaques were studied in detail, these types did not appear to emerge at random but rather in a predictable order. The evidence strongly suggests that serial passage in vitro imposed environmental pressures that sequentially selected the following viral types: Ls, Lu, Su and Ss. The data also raise a question of the possible existence of other as yet undiscerned viral types.

Virus from plaques that grew to diameters of approximately 2 mm was, on occasion, difficult to evaluate on the basis of plaque size alone. Other investigators^{18,19} have reported intermediate-plaque virus among their viral populations. On the other hand, unpublished data obtained in this laboratory and other reports¹⁷⁻¹⁹ reviewed recently by Barron and Karzon¹⁹ have shown that inhibitors in the agar influence the growth and plaque-producing ability of certain particles within a population. One explanation of this was shown by Colón et al.²⁰ to be an interaction between agar polysaccharide and susceptible virus particles of which our attenuated VEE virus was an example. In our work, virus from some 2-mm plaques, occasionally found in attenuated virus material during experiments, produced only small plaques upon passage and was attenuated for test animals. Virus from other 2-mm plaques produced plaques of equal or larger size containing virulent virus. Whether our encounters with plaques of approximately 2 mm in size resulted from the temporary alteration of environmental conditions that affected only plaque size or whether these plaques contained, in all cases, unstable intermediate-plaque virus is not presently known. The influence of the viral-inhibiting factor in agar may vary despite precaution. This could impose some degree of uncertainty in immediately recognizing the genuine intermediate-sized plaques. Evidence for the selection of intermediate-plaque virus from large-plaque preparations and results of additional work on the recognition and characterization of intermediate-plaque-forming virus are to be presented in two subsequent reports, one in preparation and one in press.⁹

LITERATURE CITED

1. Koprowski, H.; Lennette, E.H. 1946. Effect of in vitro cultivation on the pathogenicity of Venezuelan equine encephalomyelitis virus. J. Exp. Med. 84:205-210.
2. Murphy, L.C.; Blackford, V.L.; Gleiser, C.A. 1955. Study of the properties of the virus of Venezuelan equine encephalomyelitis modified by in vitro cultivation in HeLa cells. Amer. J. Vet. Res. 16:521-524.
3. Hearn, H.J., Jr. 1960. A variant of Venezuelan equine encephalomyelitis virus attenuated for mice and monkeys. J. Immunol. 84:626-629.
4. Berge, T.O.; Banks, I.S.; Tigertt, W.D. 1961. Attenuation of Venezuelan equine encephalomyelitis virus by in vitro cultivation in guinea-pig heart cells. Amer. J. Hyg. 73:209-218.
5. Mussgay, M.; Suarez, O. 1962. Studies with a pathogenic and an attenuated strain of Venezuelan equine encephalomyelitis virus and Aedes aegypti (L.) mosquitoes. Arch. Ges. Virusforsch. 12:387-392.
6. McKinney, R.W.; Berge, T.O.; Sawyer, W.B.; Tigertt, W.D.; Crozier, D. 1963. Use of an attenuated strain of Venezuelan equine encephalomyelitis virus for immunization in man. Amer. J. Trop. Med. Hyg. 12:597-603.
7. Hardy, F.M.; Hearn, H.J., Jr. 1961. The formation of plaques by two strains of Venezuelan equine encephalomyelitis virus. Amer. J. Hyg. 73:258-262.
8. Brown, A. 1963. Differences in maximum and minimum plaque-forming temperatures among selected group A arboviruses. Virology 21:362-372.
9. Heydrick, F.P.; Wachter, R.F.; Hearn, H.J., Jr. 1966. Host influence on the characteristics of Venezuelan equine encephalomyelitis virus. J. Bacteriol. In press. Also, Technical Manuscript 276. Virus and Rickettsia Division, U.S. Army Biological Laboratories, Frederick, Maryland.
10. Dulbecco, R. 1952. Production of plaques in monolayer tissue cultures by single particles of an animal virus. Proc. Nat. Acad. Sci. 38:747-752.
11. Vogt, M.; Dulbecco, R.; Wenner, H.A. 1957. Mutants of poliomyelitis viruses with reduced efficiency of plating in acid medium and reduced neuropathogenicity. Virology 4:141-155.

12. McClain, M.E.; Hackett, A.J. 1959. Biological characteristics of two plaque variants of vesicular exanthema of swine virus, type E₅₄. *Virology* 9:577-597.
13. Hsiung, G.D. 1960. Studies on variation in Coxsackie A-9 virus. *J. Immunol.* 84:285-291.
14. Hannoun, C.; Asso, J.; Ardoin, P. 1964. Mutants a petites plages du virus Sindbis. *Ann. Inst. Pasteur* 107:598-603.
15. Marshall, I.D.; Scrivani, R.P.; Reeves, W.C. 1962. Variation in the size of plaques produced in tissue culture by strains of western equine encephalitis virus. *Amer. J. Hyg.* 76:216-224.
16. Walen, K.H. 1963. Demonstration of inapparent heterogeneity in a population of an animal virus by single-burst analyses. *Virology* 20:230-234.
17. Liebhaver, H.; Takemoto, K.K. 1961. Alteration of plaque morphology of EMC virus with polycations. *Virology* 14:502-504.
18. Takemoto, K.K.; Liebhaver, H. 1961. Virus-polysaccharide interactions: I. An agar polysaccharide determining plaque morphology of EMC virus. *Virology* 14:456-462.
19. Barron, A.L.; Karzon, D.T. 1965. Studies of mutants of echovirus 6: I. Biologic and serologic characteristics. *Amer. J. Epidemiol.* 81:323-332.
20. Colón, J.I.; Idóine, J.B.; Brand, O.M.; Costlow, R.D. 1965. Mode of action of an inhibitor from agar on growth and hemagglutination of group A arboviruses. *J. Bacteriol.* 90:172-179.

Unclassified
Security Classification

| DOCUMENT CONTROL DATA - R&D | | |
|--|--|---|
| (Security classification of title, body of abstract and indexing annotation must be entered when the overall report is classified) | | |
| 1. ORIGINATING ACTIVITY (Corporate author) | | 2a. REPORT SECURITY CLASSIFICATION |
| U.S. Army Biological Center Fort Detrick, Frederick, Maryland, 21701 | | Unclassified |
| | | 2b. GROUP |
| 3. REPORT TITLE | | |
| PROPERTIES OF VENEZUELAN EQUINE ENCEPHALOMYELITIS VIRUS ACCOMPANYING ATTENUATION IN VITRO | | |
| 4. DESCRIPTIVE NOTES (Type of report and inclusive dates) | | |
| 5. AUTHOR(S) (Last name, first name, initial) | | |
| Hearn, Henry J., Jr. Soper, William T. | | |
| 6. REPORT DATE | 7a. TOTAL NO. OF PAGES | 7b. NO. OF REFS |
| June 1966 | 16 | 20 |
| 8a. CONTRACT OR GRANT NO. | 9a. ORIGINATOR'S REPORT NUMBER(S) | |
| a. PROJECT NO. 1C014501B71A | Technical Manuscript 294 | |
| c. | 9b. OTHER REPORT NO(S) (Any other numbers that may be assigned this report) | |
| d. | | |
| 10. AVAILABILITY/LIMITATION NOTICES | | |
| Qualified requesters may obtain copies of this publication from DDC. Foreign announcement and dissemination of this publication by DDC is not authorized. Release or announcement to the public is not authorized. | | |
| 11. SUPPLEMENTARY NOTES | | 12. SPONSORING MILITARY ACTIVITY |
| | | U.S. Army Biological Center Fort Detrick, Frederick, Maryland, 21701 |
| 13. ABSTRACT | | |
| <p>Virus obtained during serial plaque passage of the virulent parent egg seed (PES) strain of Venezuelan equine encephalomyelitis (VEE) virus produced only large plaques during either three serial plaque passages in chick fibroblasts or ten plaque passages in L cells and was lethal for mice by the intraperitoneal (IP) route. Virus showing these characteristics was designated the stable large-plaque (Ls) type. In contrast, virus obtained during serial plaque passage of the attenuated 9t strain in chick fibroblasts formed only very small plaques and was not lethal for mice by the IP route. Virus showing these properties was designated the stable small-plaque (Ss) type. Under other passage conditions, however, large-plaque virus that yielded about 90% large and 10% small plaques was obtained; this virus was designated the unstable large or Lu type because it differed from the Ls type, which yielded only large plaques. The Lu type continued to yield the same ratio of large to small plaques for several plaque-to-plaque passages. In addition, small-plaque virus that yielded both large and small plaques and that showed a reduced capability to infect mice was also recovered. This virus was designated the unstable small or Su type because it differed from the Ss type in its higher level of virulence and in its plaque-forming properties. Thus, based upon the properties of virulence for mice and plaque size, four viral types could be discerned. The evidence suggests that serial passage in cell culture imposed environmental pressures that sequentially selected the following viral types: Ls, Lu, Su, and Ss.</p> | | |

DD FORM 1473

Unclassified
Security Classification